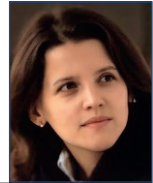


The lifestyle of the leopard gecko and the importance of ultraviolet radiation, vitamin D and calcium



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Abstract

The gecko leopard (*Eublepharis macularius*) was first described in 1854 by British herpetologist Edward Blyth. It is one of the most popular pet lizard species, due to its docile temperament, ease of maintenance and reproduction in captivity, its high longevity and small size, and its beauty and diversity of colours and patterns. Ultraviolet (UV) radiation is an important behavioural regulator. Vitamin D also regulates many fundamental physiological functions in vertebrates, mainly calcium

homeostasis. Because of the great diversity of reptile species and the wide range of environmental adaptations, it is important to know the necessities and adaptations of each species regarding UV, vitamin D and calcium requirements. This study provides an overview of the leopard gecko's lifestyle, and the importance of ultraviolet radiation, vitamin D and calcium for this species.

Key words: *behaviour; Eublepharis macularius; habitat; life; supplementation*

Leopard gecko

Eublepharis macularius was first described in 1854 by Edward Blyth, a British herpetologist (Khan, 2016). The genus *Eublepharis*, which was first described in 1827 by the British zoologist John Edward Gray, results from the combination of the words "eu", meaning "good" or "true", and "blephar", meaning "eyelid", and the main characteristic of this

genus is the presence of movable eyelids. The word "macularius" means "spotted" (Mirza et al., 2014; de Vosjoli et al., 2017). Commonly known as the leopard gecko, it is one of the most popular lizard species as a pet, mainly due to its docile temperament, ease of maintenance and reproduction in captivity, its high longevity and small size, and its beauty and di-

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versity of colours and patterns (de Vosjoli et al., 2017). There are several records of these animals living for more than 20 years in captivity, and the maximum age recorded in this species is 28 years and six months at the St. Louis Zoo (Shifter, 1988; Slavens and Slavens, 2001). From the scientific point of view, it is a species of great interest and frequently studied due to its regeneration ability (McLean et al., 2011; Delorme et al., 2012; Peacock et al., 2015; Nakashima, 2016; McDonald et al., 2018) and its thermo-dependent sexual determination and behaviour (Viets et al., 1993; Flores et al., 1994, 1995; Rhen et al., 2000; Putz et al., 2005; Pokorná et al., 2010; Huang et al., 2012, 2014).

Taxonomy

Based on the currently accepted taxonomic framework, *Eublepharis macularius* belongs to the Phylum *Chordata*, Sub-phylum *Vertebrata*, Class *Reptilia*, Order *Squamata*, Suborder *Sauria*, Family *Gekkonidae*, Subfamily *Eublepharinae*, Genus *Eublepharis* and Species *Eublepharis macularius* (Khan, 2016). They are born about 8 cm in length and in the adult stage length varies between 20 and 25 cm and weight between 45 and 60 g. Generally, males are larger than females and have 9 to 14 pre-cloacal pores arranged in an arc shape and two protuberances at the base of the tail. In the wild, colouration varies from straw yellow to pinker tones with black spots. However, due to their high popularity, and selections and crosses made by breeders, there are more than 160 mutations and combinations of different colours, patterns, and sizes (Kratochvíl and Frynta, 2002; Khan, 2016; de Vosjoli et al., 2017).

Habitat

The natural habitat is distributed across Pakistan, Iran, Afghanistan, India, and Nepal (Khan, 2016; Rawat et al.,

2019). In urbanised areas, they colonise holes and cracks in man-made structures, such as walls, roads, bridges and spaces close to underground pipes that provide the necessary moisture and shelter (Khan, 2016). They also inhabit rocky rural areas with bushes and shelter in holes and cracks in the soil, rocks, and stones. They avoid deserts and prefer humid places. They are nocturnal animals, leaving the shelter at dusk to search for food alone, and returning before dawn (Khan, 2016). In its natural habitat, in Pakistan, temperatures vary between 22-24°C in March and 40-45°C in June and July, and the relative humidity varies between 30-40%. During the monsoon season, temperatures drop to 28-33°C and humidity rises to 70-80%. Between March and June, the humidity in the shelters remains between 40 and 56%, which is ideal for these animals (Khan, 2016). In colder areas, they hibernate from September until March, while in warmer areas hibernation may be delayed until November or may be absent (Khan, 2016).

Social organisation and reproduction

Gecko colonies, before they reach sexual maturity (pre-breeding colony), are made up of several young males and females. However, when they reach sexual maturity, they become aggressive and territorial, leading to the dispersion of males, leaving only the most dominant with several females (breeding colony). The reproductive season extends from March to July and, generally, females lay two eggs per clutch (Khan, 2016). The sex determination of this species is thermo-dependent. Thus, when incubation takes place at lower temperatures (26 to 30°C) and higher (34 to 35°C) exclusively or mostly females are born, while at intermediate temperatures (31 to 33°C) mostly males are born. The incubation period is inversely proportional to the incubation

temperature, between 26°C and 32.5°C, varying between 36 and 72 days (Viets et al., 1993).

Food

Leopard geckos are mainly insectivores, feeding on beetles and grasshoppers, spiders, scorpions, and centipedes. As they age, they become opportunistic predators and attack smaller geckos, some snakes, newborn mice and bird nests (Khan, 2016). A case of cannibalism has been described in young animals (about 2–3 months old) kept in captivity (Bonke et al., 2011).

Threats in Nature

In its natural habitat, the leopard gecko is preyed on by various wild animals, such as foxes, jackals, mongooses, owls, kites, varanids and other lizards, and most snakes present in the same areas (Khan, 2016). In addition to these predators, and despite being one of the most popular pet reptiles in the world, in some areas of its natural habitat, local people consider this species to be a poisonous animal, believing it is related to the snakes *Naja naja karachiensis* and that its bite liquefies the victim's body, causing instantaneous death. For this reason, many of these native peoples kill geckos upon sight (Khan, 2016). There is currently no information available about the conservation status of this species in its natural habitat (IUCN Red List, 2020).

Captivity

According to Wright (2011, as cited by Boyer, 2013), the leopard gecko is the second most common reptile species in veterinary practice (13% of reptile cases). Most cases result from poor conditions in captivity. Some of the most common pathologies are nutritional pathologies (nutritional hyperparathyroidism, hypovitaminosis A), pathologies associated

with deficiencies in skin shedding (dys-eecdysis), pathologies of the gastrointestinal tract (gastrointestinal obstructions and other pathologies in the gastrointestinal system, often associated with cryptosporidiosis), pathologies of the reproductive system, trauma (including tail autotomy), ocular pathologies and subcutaneous abscesses (Boyer, 2013; Miles, 2017). When kept in captivity, as terrestrial animals, the terrarium should be horizontal and as large as possible. However, the minimum acceptable volume of a terrarium to keep a specimen is about 38 litres, with the length of the terrarium at least twice the length of the animal and the width at least equal to the length of the animal (Bartlett, 1995; Wilkinson, 2015; deVosjoli, 2017). Various artificial or natural materials can be used as substrate. Artificial substrates, such as kitchen paper or newspaper sheets, are advantageous due to their low price and easy cleaning, however, they are less aesthetically appealing than natural substrates. If natural substrates are chosen, the risk of intestinal impaction must be considered and substrates that are not particulate or easily digestible must be chosen (Wilkinson, 2015; de Vosjoli et al., 2017). Leopard Geckos have an average ideal body temperature of 28.2±0.6°C, increasing significantly throughout the day (Angilletta et al., 1999). A thermal gradient must be maintained in the terrarium, to allow the animals behavioural thermoregulation, optimising metabolism and stimulating growth (Autumn, 1995). Thus, the animal must be provided with an area of the terrarium with a temperature between 29 and 33°C, and a colder zone at about 24°C. During the night the temperature can drop to 18° if a warm spot is maintained in the terrarium. This can be done, for example, by placing a heating mat under the terrarium that occupies 25 to 35%

of its area (de Vosjoli et al., 2017). It is also important to maintain an adequate relative humidity gradient to facilitate ecdysis or oviposition and to prevent chronic dehydration. The most frequently used method consists of placing a substrate that can maintain humidity (for example a substrate of Sphagnum moss) in a plastic box and making an opening in the lid so that the animals can get in and out (Rossi, 2019). Feeding must be based on insects or other live invertebrates, as these provide the animals with important environmental enrichment and can prolong the feeding period. However, most foods available on the market are nutritionally unbalanced if not supplemented in an appropriate way. For example, insects in the larval stage generally have a disproportionately higher amount of fat than the amounts of other nutrients, most insects are poor sources of calcium, and annelids, despite having less fat and high calcium values, have the same very variable nutritional contents, depending on the composition of the substrate where they are kept. The common practice of spraying or dipping insects with calcium supplements can provide inconstant or insufficient levels of calcium, can change their palatability and, if the insect is not consumed immediately, they can lose part or all the supplement (Bernard et al., 1997; Li et al., 2009). To combat this problem, there are already commercial diets rich in calcium for crickets that can be used to feed other insects. In addition, calcium supplements such as calcium carbonate can be added to the insect diet to increase its calcium content (Zwart et al., 1979; Strzelewicz et al., 1985; Allen et al., 1989; Finke, 2013).

Ultraviolet radiation

Ultraviolet (UV) radiation is a natural component of sunlight that is subdivid-

ed into three groups: the portion of UV radiation with the shortest wavelength (100–290 nm) is UVC radiation, the portion with an intermediate wavelength (290–320 nm) is UVB radiation, and the portion with the longest wavelength (320–400 nm) is UVA radiation (Adkins et al., 2003; Baines et al., 2016). UVB radiation enables the conversion of 7-dehydrocholesterol present in the skin into pre-vitamin D₃ which, by thermal action, is converted into cholecalciferol (vitamin D₃) (Baines et al., 2016). Pre-vitamin D₃ produced in excess is converted into inert photoproducts, lumisterol and tachysterol, by the action of UVB radiation and low-wavelength UVA radiation, *i.e.*, at wavelengths between 290 and 335 nm. This is a self-limiting process that prevents overproduction of vitamin D₃ (MacLaughlin et al., 1982, Webb et al., 1989). UVC radiation, which is normally completely filtered by the atmosphere, causes cell damage and therefore is not necessary and should not be added as an artificial source of radiation (Adkins et al., 2003).

Transmission of ultraviolet radiation

The skin acts as a barrier between the organism and the environment, protecting it from mechanical abrasion, microorganisms, water loss and UV radiation. Reptile skin, like the skin of many other vertebrates, has two main layers: the epidermis and the dermis (Subramaniam et al., 2018). The epidermis consists of a keratinised stratified epithelium and in reptiles of the order Squamata, which includes the leopard gecko, it is divided into several sub-layers: Oberhäutchen layer, β -keratin layer, α -keratin layer, intermediate zone, and germinal layer (basal stratum). The combination of the Oberhäutchen, β -keratin and α -keratin layers constitute the stratum corneum

(Subramaniam et al., 2018). The Oberhäutchen layer is the most superficial and hardest layer of the epidermis and is made up of dead flattened and anucleated cells, with keratin-rich cytoplasm. The β -keratin layers are composed of cells undergoing keratinisation, through the production of two types of keratin: α keratin, which is flexible, and β keratin, which is strong and hard, and only exists in reptiles. In this way, these layers become a hard and protective layer. The epidermis is in constant renewal, with the cells of the stratum germinativum as progenitor cells. After proliferation, the new keratinocytes migrate through the intermediate zone, until they reach the fully differentiated stratum corneum. Over time, these keratinocytes desquamate and will be replaced by the new generation of keratinocytes (Maderson et al., 1998; O'Malley, 2005; Subramaniam et al., 2018). Melanin, which is the pigment responsible for brown coloration, is found in epidermal cells and effectively absorbs UV radiation at wavelengths between 200 and 700 nm, which includes the UVB radiation responsible for the cutaneous production of vitamin D₃ (Jablonski et al., 2000). Thus, the amount of melanin present in the epidermis will determine the amount of radiation that reaches the deeper layers of the skin (O'Malley, 2005; Baines et al., 2006; Rutland et al., 2019). It is in the membrane of the deepest cells of the epidermis that, due to the incidence of UV radiation, 7-dehydrocholesterol is converted into pre-vitamin D₃ (Holick, 2004; Junqueira and Carneiro, 2013). UV radiation, before reaching these cells, crosses the most superficial layers of the epidermis. The epidermis of reptiles varies greatly from species to species both in thickness and in pigmentation. Microscopic observation of the skin of a green iguana (*Iguana iguana*), for example, has

shown that it has a much higher level of keratinisation when compared to the skin of leopard geckos. In addition, the green iguana has the most pronounced β -keratin layer and very few melanocytes. This larger layer of keratin prevents the transmission of radiation through the epidermis and, consequently, when compared to the percentage of UV radiation passing through the skin of the two species, about four times more radiation will pass through the skin of the leopard gecko than the green iguana. The skin of lizard species that are exposed to higher intensities of radiation in their habitat shows a tendency to block more radiation, when compared with the skin of nocturnal, crepuscular species or those in areas with lower radiation intensity (Baines et al., 2006; Rutland et al., 2019). The dermis is the deepest layer of the skin, consisting of dense connective tissue, including adipocytes, blood vessels, lymphatic vessels, nerves, inflammatory cells and two types of pigment cells: xanthophores (yellow pigment) and melanophores (black pigment) (O'Malley, 2005; Szydlowski et al., 2015; Rutland et al., 2019). Season is also an important factor in transmitting radiation useful to produce vitamin D₃. A study performed in humans demonstrated that during June and July, the efficiency of vitamin D₃ production reached its peak, gradually decreasing after August, and ceasing almost completely between November and March (Webb et al., 1988). Latitude is another factor to consider, since it influences the zenith angle and, consequently, the intensity and period of exposure to solar radiation. In places closer to the poles, vitamin D₃ production ceases for six months of the year, while in places closer to the equator its synthesis occurs uninterrupted throughout the year (Wacker and Holick, 2013a; Holick, 2018).

Benefits of ultraviolet radiation

Exposure to UV radiation brings several advantages to the organism (Juzeniene et al., 2012). Exposure to this radiation results in the activation of p53 which, in turn, regulates the expression of the gene encoding pro-opiomelanocortin. This is a polypeptide precursor, whose cleavage originates several bioactive products, among them, the melanocyte-stimulating hormone- α (α -MSH) and β -endorphin (Oren et al., 2007; Juzeniene et al., 2012). α -MSH is responsible for activating the production of melanin, which effectively absorbs UV radiation, protecting the skin from future exposure to this radiation (Catania et al., 2010). β -endorphin is the most abundant endogenous neurotransmitter in the blood and has analgesic effects and produces a sense of well-being (Bernard et al., 1991; Juzeniene et al., 2012; Fell et al., 2014; Veleva et al., 2018). In addition, UV radiation also promotes hyperkeratosis, thus providing greater protection (Bulat et al., 2011). UV radiation can also be used in the treatment and relief of symptoms of certain diseases. Heliotherapy (treatment with the use of sunlight) was already used in ancient Greece, Egypt and Rome for the treatment of various skin pathologies (Roelandts et al., 2002). There is evidence to suggest that exposure to UV radiation can decrease short-term pain in patients with fibromyalgia (Taylor et al., 2009). Access to this type of radiation is important to stimulate the natural behaviour of many species in captivity (Honkavaara et al., 2002). Several species regulate the period of exposure to radiation according to their physiological needs (Manning et al., 1997; Ferguson et al., 2003; Karsten et al., 2009; Oonincx et al., 2010).

Adverse effects of ultraviolet radiation

When exposure to UV radiation is excessive, several adverse effects may arise, mainly on the eyes and skin, both in cases of acute exposure and long-term exposure (Hathaway et al., 2016). In animals, there are reports of photodermatitis and keratoconjunctivitis in royal pythons and only photodermatitis in blue-tongued lizards, due to the long-term use of a UV lamp with excessive intensity. Other reptiles exposed to radiation with the same characteristics showed identical pathologies: a crested gecko (*Rhacodactylus ciliatus*), an albino Burmese python (*Python molurus bivittatus*), a tree boa of the genus *Candoia*, a bearded dragon (*Pogona vitticeps*), a snake of the species *Lampropeltis triangulum hondurensis*, and two Kenyan sand boas (*Érix colubrinus colubrinus*) (Gardiner et al., 2009). A study carried out on budgerigars (*Melopsittacus undulatus*) reported several adverse effects after exposure to UV radiation of different intensities. At lower intensities, effects such as weight loss, increased concentration of corticosterone in the blood and skin erythema were observed. At higher radiation intensities, animals developed corneal photokeratitis (Lupu et al., 2013). As in humans, skin cancers in captive reptiles may be related to exposure to UV radiation. It is possible that there is a relationship between radiation exposure and the incidence of squamous cell carcinomas and pigment cell neoplasms. A study with 69 reptiles suggested that heliophilic species have a higher incidence of this type of neoplasms when compared to nocturnal species. The same authors concluded, in another study, that around 60% of animals with pigment cell neoplasms had access to artificial UV radiation. However, these results are not conclusive and further studies are needed to understand this relationship in reptiles (Heckers et al., 2012, 2014).

Vitamin D and Calcium

At the end of the 19th century, after the industrial revolution, there was an outbreak of a disease characterised by skeletal deformities in children, known as rickets. Sir Edward Mellanby, to respond to this new outbreak, successfully used cod liver oil and the anti-rachitic properties of this substance were initially attributed to vitamin A (DeLuca et al., 2018). Vitamin D was discovered in 1922 by Elmer McCollum. After destroying the vitamin A present in cod liver oil, McCollum observed that it maintained its ability to cure rickets and concluded that the therapeutic properties of cod liver oil were not due to vitamin A, but to another substance which he called vitamin D (McCollum et al., 1922). Later, in 1925, Steenbock discovered that UV radiation had the ability to activate an inactive substance, giving rise to vitamin D (Steenbock et al., 1925).

Acquisition of vitamin D

Vitamin D can be acquired in two ways: through food and subsequent absorption in the gastrointestinal tract or through synthesis in the skin after exposure to UVB radiation (Hossein-Nezhad and Holick, 2013). In nature, vitamin D is available as ergocalciferol (vitamin D₂), whose precursor is ergosterol (provitamin D₂), or as cholecalciferol (vitamin D₃), whose precursor is 7-dehydrocholesterol (7-DHC) (pro-vitamin D₃) (Chen et al., 2010; Polzonetti et al., 2020). The difference between the two precursors is in the presence of an extra methyl group on carbon 24 of ergosterol (Chen et al., 2010; Engelking, 2015). Most authors consider that ergocalciferol (vitamin D₂) is found mainly in plants and fungi, and cholecalciferol (vitamin D₃) is found in products of animal origin (Chen et al., 2016; Polzonetti et al., 2020), though there is no

consensus, since other authors consider that ergocalciferol is found only in fungi, while cholecalciferol is found in both animals and plants, albeit in small amounts (Wacker and Holick, 2013b; Göring et al., 2018). Vitamin D₃ can be obtained by eating foods that naturally contain this vitamin (cod liver oil, fatty fish such as salmon and tuna), mushrooms or plants exposed to UV radiation, foods fortified with vitamin D or through dietary supplements (Hossein-Nezhad and Holick, 2013). Alternatively, vitamin D₃ can be synthesised in the skin through exposure to UVB radiation. The skin of most vertebrates contains the precursor of vitamin D₃ (7-DHC) in the membranes of the deepest cells of the epidermis (Holick et al., 2014). Thus, when the skin is exposed to radiation with a wavelength between 290 and 320 nm (UVB radiation), 7-DHC is converted into pre-vitamin D₃ (Bunker et al., 1940; Chen et al., 2010; Watson, 2014). The energy of the photons, when incident on 7-DHC, causes a break in the bond between carbon 9 and carbon 10 of the molecule and an isomerization of the 5,7-diene, forming pre-vitamin D₃. This molecule is unstable and rapidly undergoes a rotation on the bond between carbon 5 and carbon 6 and a rearrangement of its hydrogen atoms, through a thermo-dependent process, giving rise to vitamin D₃ or cholecalciferol (Holick et al., 1995; Chen et al., 2010; Holick, 2018). After being produced in the skin, vitamin D₃ is transported to the liver where it undergoes hydroxylation at carbon 25, mediated by the enzyme 25-hydroxylase. 25-hydroxycholecalciferol, 25-hydroxyvitamin D₃ or calcidiol (25(OH)D₃) is the most abundant vitamin D metabolite in the blood and is not biologically active; however, it plays a very important role as a reserve form of this vitamin. In addition to its presence in the blood, it is stored in

different places such as the liver, adipose tissue, and skeletal muscle tissue (Dahlback et al., 1988; Cline, 2012; Holick, 2014; Jones et al., 2018). To become biologically active, this form of vitamin D has to undergo a second hydroxylation, this time at carbon 1. This hydroxylation is mediated by the enzyme 1α -hydroxylase and gives rise to 1,25-dihydroxyvitamin D₃ (1,25(OH)₂D₃), 1,25-dihydroxycholecalciferol or calcitriol, the active hormonal form of vitamin D₃ (Zehnder et al., 2002; Cline, 2012; Holick, 2014). This reaction occurs mainly in the kidneys (Zehnder et al., 2001, 2002; Larner et al., 2018). Vitamin D₂ or ergocalciferol can only be obtained through food. Like vitamin D₃, it undergoes two hydroxylations: the first in the liver and the second in different parts of the body, giving rise to 25-hydroxyergocalciferol or ercalcidiol and 1,25-dihydroxyergocalciferol or ercalcitriol, respectively (Holick, 2014; Dereje et al., 2017).

Vitamin D transport

Lipophilic steroid molecules, such as vitamin D, bind to plasma proteins in their blood transport. The main transport protein for vitamin D and its metabolites is vitamin D binding protein. This protein is responsible for about 80% of transport, while proteins such as albumin and lipoproteins, which are more abundant in the blood, have a less important role in this transport (Bouillon and Pauwels, 2017). Vitamin D binding protein, despite transporting all vitamin D metabolites, has a higher affinity for transporting calcidiol and a lower affinity for transporting calcitriol or vitamin D itself. In most species, this protein has a higher affinity for vitamin D₃ metabolites than for vitamin D₂ metabolites. In birds, the affinity of vitamin D binding protein for transporting vitamin D₃ metabolites is three to ten

times greater than the affinity for transporting vitamin D₂ metabolites (Haddad et al., 1976; Bouillon et al., 1980; Marx et al., 1989; Bouillon and Pauwels, 2017).

Vitamin D functions

Vitamin D's main function is to maintain blood calcium and phosphorus levels within the values considered physiological and necessary for various metabolic functions, the regulation of gene expression and the success of bone metabolism (Fleet, 2018). It is estimated that vitamin D influences between 200 and 2000 genes, directly or indirectly, suggesting that an increase in vitamin D values can regulate the expression of genes related to oncological pathologies, autoimmune pathologies, and cardiovascular diseases which, in turn, are associated with a deficiency of this vitamin (Nagpal et al., 2005; Hossein-Nezhad and Holick, 2013). Vitamin D plays, together with parathyroid hormone (PTH), an important role in bone remodelling, affecting the metabolism of calcium and phosphorus (Carmeliet, 2018). When the calcium balance is normal or positive, vitamin D and PTH are responsible for keeping calcium levels within their normal range. However, when the calcium balance is negative, the parathyroid is stimulated, increasing PTH secretion. In turn, PTH stimulates the production of the active form of vitamin D, 1,25(OH)₂D₃, through increased renal expression of 1α -hydroxylase. These two hormones promote bone remodelling, as they stimulate osteoblasts to produce cytokines that accelerate osteoclast maturation and, consequently, the release of calcium and phosphorus into circulation (Suda et al., 2015; Carmeliet, 2018; Nakamichi et al., 2018; Charoenngam et al., 2019). 1,25(OH)₂D₃ stimulates the secretion of fibroblast growth factor (FGF23) in osteocytes. This hormone, unlike PTH,

suppresses the activity of 1α -hydroxylase in the kidneys, reducing its production. The activity of the enzyme is also suppressed by the active form of vitamin D itself, which further suppresses PTH secretion in the parathyroid through its interaction with vitamin D receptors in the glands (Suda et al., 2015; Carmeliet, 2018; Nakamichi et al., 2018; Charoenngam et al., 2019). The increase in the concentration of $1,25(\text{OH})_2\text{D}_3$ also promotes the expression of 24-hydroxylase, the enzyme responsible for the inactivation of vitamin D through the hydroxylation of carbon 24, culminating in the formation of calcitroic acid. The expression of this enzyme is suppressed by PTH (Tanaka and DeLuca, 1981; Holick, 2014). Both PTH and vitamin D stimulate renal calcium reabsorption. On the other hand, the increase in phosphorus blood concentrations stimulate the secretion of PTH and FGF23 which, consequently, increase its renal excretion. In addition, vitamin D plays a very important role in the absorption of calcium at the intestinal level, through two different mechanisms: through transcellular transport, active and saturable, which occurs mainly in the proximal small intestine (duodenum and jejunum), or through passive and non-saturable paracellular transport, which occurs with the same intensity throughout the entire intestine (Pansu, 1981). Vitamin D appears to play an active role in these two mechanisms. Without it, the body is only able to absorb 10 to 15% of the ingested calcium. However, the interaction of the active form of vitamin D with vitamin D receptors in the intestine can increase the efficiency of absorption to much higher values, 30% to 40% (Holick, 2007). In animals deficient in this vitamin, absorption efficiency can decrease by more than 75% (Pansu et al., 1983).

Photoisomerisation and photodegradation of vitamin D

The body has several vitamin D regulation mechanisms that prevent its excessive production (Chen, 2010). Pre-vitamin D produced in the skin can be converted into vitamin D_3 or other photoproducts such as tachysterol and lumisterol (Holick et al., 1981; MacLaughlin et al., 1982). In addition, the vitamin D_3 produced and accumulated in the skin is sensitive to radiation and is degraded when exposed, creating various photoproducts. A study showed that solar radiation in summer in Boston can degrade more than 50%, 75% and 95% after 30 minutes, one hour and three hours of exposure, respectively. The main photoproducts found were suprasterol 1, suprasterol 2 and 5,6-transvitamin D_3 (analogous molecule of vitamin D_3) (Webb et al., 1989). Thus, prolonged exposure to UV radiation does not continuously increase the plasma concentration of $25(\text{OH})\text{D}_3$ (Watson, 2014).

Vitamin D intoxication

Vitamin D in high doses can be toxic (O'Malley, 2008). However, there are no reported cases of pathologies derived from excess vitamin D caused by excessive exposure to radiation. This happens since both pre-vitamin D_3 and vitamin D_3 itself are susceptible to radiation, giving rise to various products that are inert or inactive in calcium homeostasis (Baines et al., 2006; Chen et al., 2010; Watson, 2014). Vitamin D intoxication caused by a natural diet is rare, and most cases described in reptiles are derived from an overdose of supplementation or an inadequate diet, as is the case of reptiles fed with dog or cat food containing high concentrations of vitamin D (Wallach, 1996; O'Malley, 2008; Penning, 2012; Boyer and Scott, 2019). In addition, intoxication

can be caused acutely by the ingestion of cholecalciferol-based rodenticides or topical drugs for psoriasis, although this hypothesis is unlikely in captive reptiles (Cline, 2012; Dee and Hovda, 2012). Hypervitaminosis D can be difficult to diagnose, mainly due to the slow onset of clinical signs and, therefore, the prognosis can be very guarded (Cline, 2012). Excess vitamin D causes hypercalcemia and, consequently, mineralisation of the soft tissues of the gastrointestinal tract, muscles, kidneys, lungs, heart, and large vessels, which can usually be observed on radiographs or ultrasounds, and in the long term can cause bone malformations. These mineralisations can cause pain and organ dysfunction or failure (Wallach, 1996; Raiti and Garner, 2006; O'Malley, 2008; Cline, 2012; Penning, 2012; Watson, 2014; Boskey, 2018; Boyer and Scott, 2019). Currently there are no guidelines for the treatment of these pathologies in reptiles and therefore the treatment recommended for others is followed, based on glucocorticoids, calcitonin and, according to more recent studies, pamidronate (O'Malley, 2008; Boyer and Scott, 2019).

Vitamin D deficiency

In reptiles, the main consequence of hypovitaminosis D is metabolic bone disease (MBD) caused by nutritional secondary hyperparathyroidism (Watson, 2014). MBD is a term used to describe a set of diseases that affect the integrity and function of bones (Carmel and Johnson, 2017). The most common MBD in reptiles is secondary nutritional hyperparathyroidism (NSHP), whose main causes are prolonged calcium or vitamin D deficiency, an imbalance in the calcium-phosphorus ratio in the diet, inadequate exposure to UV radiation or the lack of adequate temperatures in the facilities (Carmel and Johnson, 2017; Lock, 2017). MBD is more

common in turtles and lizards, as many of these animals have a diet with lower concentrations of vitamin D, while snakes have greater access to vitamin D through their diet (Carmel & Johnson, 2017). Decreased blood calcium or vitamin D concentrations and increased blood phosphorus concentrations increase PTH secretion, which in turn stimulates bone resorption. When deficiencies are prolonged, bone tissue destroyed during bone resorption is not recovered, causing a decrease in bone density (osteopenia) and weakening of the bones (Lock, 2017). The most common clinical signs include deviations in the spine and tail, swelling and stiffness of the hind limbs, bone fractures, bowed and soft long bones, deformed carapaces in turtles, difficulty in lifting the body or tail, swollen and soft jaw ("rubber jaw"), tremors, paresis or paralysis of the limbs, and lethargy (O'Malley, 2008; Penning, 2012; Lock, 2017). The recommended treatment includes supplementation with calcium and vitamin D, nutritional support, improvement of the conditions of the facilities, analgesia, and calcitonin (it is not recommended to administer calcitonin in animals with hypocalcemia or with clinical neurological signs) (Lock, 2017; Boyer and Scott, 2019). Prognosis varies with the severity of clinical signs, but lizards and crocodiles respond better to treatment than chelonians. The prognosis is good in cases without a visible decrease in bone radiopacity, whereas animals with paresis, paralysis or constipation have a worse prognosis. Good nutrition with adequate calcium supplementation and UV radiation is essential (Boyer and Scott, 2019).

Ultraviolet radiation and vitamin D in different species

Due to the great diversity of species and diverse environmental adaptations,

studies have been carried out to understand the individual needs of each species (Vergneau-Grosset et al., 2020). Most vertebrate animals contain 7-dehydrocholesterol in the skin in sufficient concentrations to produce significant amounts of vitamin D₃ when exposed to UV radiation (Holick et al., 2014). A study carried out on leopard geckos (*Eublepharis macularius*) demonstrated that these animals, when exposed to short periods of UV radiation (2 h), can significantly increase the concentration of 25-hydroxyvitamin D₃. These data once again demonstrate that even nocturnal species can produce vitamin D₃ when exposed to UV radiation, even for short periods (Gould et al., 2018). Indeed, several species with less opportunity for exposure to sunlight have been shown to have evolved an adaptation mechanism that allows them to synthesise vitamin D₃ more efficiently (Carman et al., 2000; Baines et al., 2006; Ferguson et al., 2015; Rutland et al., 2019). One study compared the 7-dehydrocholesterol conversion capacity of the Texas spiny lizard (*Sceloporus olivaceus*) and the gecko (*Hemidactylus turcicus*). Though the former species is diurnal, and the latter is nocturnal, the nocturnal species was found to have a greater ability to convert 7-dehydrocholesterol (Carman et al., 2000).

Conclusions

The leopard gecko is one of the most popular pet lizard species, mostly due to its docile temperament, ease of maintenance and reproduction in captivity, its high longevity and small size, its beauty and diversity of colours and patterns. Since this species is frequently maintained in captivity, it is important to clearly understand the importance and its needs in terms of UV radiation, vitamin D and calcium for its normal behav-

iour and maintenance of fundamental physiological functions.

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References

1. ADKINS, E., T. DRIGGERS, G. FERGUSON, W. GEHRMANN, Z. GYIMESI, E. MAY, M. OGLE, T. OWENS and E. KLAPHAKE (2003): Ultraviolet Light and Reptiles, Amphibians. *J. Herpetol. Med. Surg.* 13, 27-37. 10.5818/1529-9651.13.4.27.
2. ALLEN, M. E., T. C. CHEN, M. F. HOLICK and E. MERKEL (1999): Evaluation of Vitamin D Status in the Green Iguana (*Iguana iguana*): Oral Administration vs UVB Exposure. *Biol. Effects of Light* 1998, 99-101. 10.1007/978-1-4615-5051-8_17.
3. ALLEN, M. and O. OFTEDAL (1989): Dietary Manipulation of the Calcium Content of Feed Crickets. *Allen, Mary E. and Ofstedal, Olav T.*, 20, 26.
4. ANGILLETTA, M. J., L. G. MONTGOMERY and Y. L. WERNER (1999): Temperature preference in geckos: Diel variation in juveniles and adults. *Herpetologica* 55, 212-222.
5. AUTUMN, K. and D. F. NARDO (1995): Behavioral Thermoregulation Increases Growth Rate in a Nocturnal Lizard. *J. Herpetol.* 29, 157-162.
6. BAINES, F., A. BEVERIDGE, R. HITCH and R. LANE (2006): The Transmission of Ultraviolet Light through Reptile Skin Shed. <http://uvguide.co.uk/skintests.htm>.
7. BAINES, F., J. CHATTELL, J. DALE, D. GARRICK, I. GILL, M. GOETZ, T. SKELTON and M. SWATMAN (2016): How much UV-B does my reptile need? The UV-Tool, a guide to the selection of UV lighting for reptiles and amphibians in captivity. *J. Zoo Aquar. Res.* 4, 42-63.
8. BARTLETT, R. D. and P. P. BARTLETT (1995): *Geckos: A Complete Pet Owner's Manual*. In *Geckos: A Complete Pet Owner's Manual* (1st ed.). Barron's Educational Series, Inc.
9. BERNARD, J., O. T. OFTEDAL, P. S. BARBOZA, C. E. MATHIAS, M. E. ALLEN, S. B. CITINO, D. E. ULLREY and R. J. MONTALI (1991): Response Of Vitamin D-Deficient Green Iguanas (*Iguana Iguana*) To Artificial Ultraviolet Light. *Am. Assoc. Zoo Vet.* 1-4.

10. BERNARD, J. B. and M. E. ALLEN (1997): Feeding captive insectivorous animals: nutritional aspects of insects as food. *Anim. Sci.* 25, 1-7.
11. BONKE, R., W. BÖHME, K. OPIELA and D. RÖDDER (2011): A remarkable case of cannibalism in juvenile leopard geckos, *Eublepharis macularius* (Blyth, 1854) (Squamata: Eublepharidae). *Herpetol. Notes* 4, 211-212.
12. BOSKEY, A. L. (2018): Mineralization in Mammals. In: Feldman, D., J. W. Pike, R. Bouillon, E. Giovannucci, D. Goltzman, and M. Hewison (Eds.), *Vitamin D: Fourth Edition: Vol. 1: Biochem* (4th ed., pp. 383-403). Elsevier Inc. 10.1016/B978-0-12-809965-0.00023-9.
13. BOUILLON, R., H. VAN BAELEN, B. KENG TAN and P. DE MOOR (1980): The isolation and characterization of the 25-hydroxyvitamin D-binding protein from chick serum. *J. Biol. Chem.* 255, 10925-10930.
14. BOUILLON, R., S. PAUWELS (2017): The Vitamin D-Binding Protein. In: Feldman, D., J. W. Pike, R. Bouillon, E. Giovannucci, D. Goltzman, and M. Hewison (Eds.), *Vitamin D: Fourth Edition: Vol. 1: Biochem* (4th ed., pp. 97-155). Elsevier Inc. 10.1016/B978-0-12-809965-0.00007-0.
15. BOYER, T., M. GARNER, D. REAVILL and Z. STEFFES (2013): Common problems of leopard geckos (*Eublepharis macularius*). *Ass. Rept. Amph. Vet.* 117-125.
16. BOYER, T. H. and P. W. SCOTT (2019): Nutritional Diseases. In: Divers, S. J. and S. J. Stahl (Eds.), *Mader's Reptile and Amphibian Medicine and Surgery* (3rd ed., pp. 932-950). Elsevier Inc.
17. BULAT, V., M. SITUM, I. DEDIOL, I. LJUBIČIĆ and L. BRADIĆ (2011): The mechanisms of action of phototherapy in the treatment of the most common dermatoses. *Coll. Antropol.* 35 Suppl 2, 147-151. <http://www.ncbi.nlm.nih.gov/pubmed/22220423>.
18. BUNKER, J. W. M., R. S. HARRIS and L. M. MOSHER (1940): Relative Efficiency of Active Wave Lengths of Ultraviolet in Activation of 7-Dehydrocholesterol. *J. Am. Chem. Soc.* 62, 508-511. 10.1021/ja01860a013.
19. CARMAN, E. N., G. W. FERGUSON, W. H. GEHRMANN, T. C. CHEN and M. F. HOLICK (2000): Photobiosynthetic Opportunity and Ability for UV-B Generated Vitamin D Synthesis in Free-Living House Geckos (*Hemidactylus turcicus*) and Texas Spiny Lizards (*Sceloporus olivaceous*). *Copeia*, 2000, 245-250. 10.1643/0045-8511(2000)2000[0245:POAAFU]2.0.CO;2.
20. CARMEL, B. and R. JOHNSON (2017): Nutritional and metabolic diseases. *Rep. Med. Surg. Clin. Pract.* 3, 185-195. <https://doi.org/10.1002/978118977705.ch15>.
21. CARMELIET, G. (2018): Vitamin D and Bone: An Integrated Approach. In: D. Feldman, J. W. Pike, R. Bouillon, E. Giovannucci, D. Goltzman, and M. Hewison (Eds.), *Vitamin D: Fourth Edition: Vol. 1: Biochem* (4th ed., pp. 419-433). Elsevier Inc. <https://doi.org/10.1016/B978-0-12-809965-0.00025-2>.
22. CATANIA, A., C. LONATI, A. SORDI, A. CARLIN, P. LEONARDI and S. GATTI (2010): The melanocortin system in control of inflammation. *Sci. World J.* 10, 1840-1853. 10.1100/tsw.2010.173.
23. CHAROENNGAM, N., A. SHIRVANI and M. F. HOLICK (2019): Vitamin D for skeletal and non-skeletal health: What we should know. *J. Clin. Orthop. Trauma.* 10, 1082-1093. 10.1016/j.jcot.2019.07.004.
24. CHEN, T., Z. LU and M. F. HOLICK (2010): Photobiology of Vitamin D. In: Holick, M. F. (Ed.), *Vitamin D. Mol. Biol. Clin. Appl.* 25, 175-187. 10.1007/978-1-60327-303-9.
25. CLINE, J. (2012): Calcium and Vitamin D Metabolism, Deficiency, and Excess. *Top. Companion Anim. Med.* 27, 159-164. 10.1053/j.tcam.2012.09.004.
26. DAHLBACK, H. and K. WIKVALL (1988): 25-Hydroxylation of vitamin D3 by a cytochrome P-450 from rabbit liver mitochondria. *Biochem. J.* 252, 207-213. 10.1042/bj2520207.
27. DE LUCA, H. F. (2018): Historical Overview of Vitamin D. In: Feldman, D., J. W. Pike, R. Bouillon, E. Giovannucci, D. Goltzman, and M. Hewison (Eds.), *Vitamin D: Fourth Edition: Vol. 1: Biochem* (4th ed., pp. 3-12). Elsevier Inc. 10.1016/B978-0-12-809965-0.00001-X.
28. DE LUCA, D. A., E. A. ZAMBRANO, R. L. GALIMBERTI and P. A. ENZ (2018): The Effectiveness of a Twice-weekly Narrowband Ultraviolet B Phototherapy Schedule in Early-stage Mycosis Fungoides in a Cohort of 18 Argentinian Patients. *Act. Dermo-Sifiliog.* 109, 922-924. 10.1016/j.adengl.2018.07.015.
29. DEVOSJOLI, P., T. MAZORLIG, R. KLINGENBERG, R. TREMPER and B. E. VIETS (2017): *The Leopard Gecko Manual: Expert Advice for Keeping and Caring for a Healthy Leopard Gecko* (2nd ed.). Fox Chapel Publishers International Ltd.
30. DEE, T. and L. R. HOVDA (2012): Cholecalciferol rodenticide toxicosis. *Vet. Techn.* 33, 3-6.
31. DELORME, S. L., I. M. LUNGU and M. K. VICKARYOUS (2012): Scar-Free Wound Healing and Regeneration Following Tail Loss in the Leopard Gecko, *Eublepharis macularius*. *Anat. Rec.* 295, 1575-1595. 10.1002/ar.22490.
32. DEREJE, S., I. MURADOV, S. NAZZAL and T. NGUYEN (2017): Cholecalciferol (D3) Versus Ergocalciferol (D2) in Older Adults. *Consult. Pharm.* 32, 337-339.
33. ENGELKING, L. R. (2015): Vitamin D. In *Textbook of Veterinary Physiological Chemistry* (3rd ed., pp. 288-293). Elsevier Inc. 10.1016/B978-0-12-391909-0.50045-1.
34. FEILY, A., E. RAFFEIE, S. RASSAI and M. RAMIREZ-FORT (2014): Adjuvant narrow band UVB improves the efficacy of oral azithromycin for the treatment of moderate to severe inflammatory facial acne vulgaris. *J. Cutan. Aesthet. Surg.* 7, 151. 10.4103/0974-2077.146664.

35. FELL, G. L., K. C. ROBINSON, J. MAO, C. J. WOOLF and D. E. FISHER (2014): Skin β -endorphin mediates addiction to UV light. *Cell* 157, 1527-1534. 10.1016/j.cell.2014.04.032.
36. FERGUSON, G. W., W. H. GEHRMANN, K. A. BRADLEY, B. LAWRENCE, R. HARTDEGEN, T. STORMS, T. CHEN and M. F. HOLICK (2015): Summer and Winter Seasonal Changes in Vitamin D Status of Captive Rhinoceros Iguanas (*Cyclura cornuta*). *J. Herpetol. Med. Surg.* 25, 128-136.
37. FERGUSON, G. W., W. H. GEHRMANN, K. B. KARSTEN, S. H. HAMMACK, M. MCRAE, T. C. CHEN, N. P. LUNG and M. F. HOLICK (2003): Do panther chameleons bask to regulate endogenous vitamin D3 production? *Physiol. Biochem. Zool.* 76, 52-59. 10.1086/374276.
38. FINKE, M. D. and D. OONINCX (2013): Insects as Food for Insectivores. In *Mass Production of Beneficial Organisms: Invertebrates and Entomopathogens*. Elsevier. 10.1016/B978-0-12-391453-8.00017-0.
39. FLEET, J. C. (2018): Regulation of Intestinal Calcium and Phosphate Absorption. In: Feldman, D., J. W. Pike, R. Bouillon, E. Giovannucci, D. Goltzman, and M. Hewison (Eds.), *Vitamin D: Fourth Edition: Vol. 1: Biochem (4th ed., pp. 329-342)*. Elsevier Inc. 10.1016/B978-0-12-809965-0.00020-3.
40. FLORES, D. L. and D. CREWS (1995): Effect of Hormonal Manipulation on Sociosexual Behavior in Adult Female Leopard Geckos (*Eublepharis macularius*), a Species with Temperature-Dependent Sex Determination. *Horm. Behav.* 29, 458-473. 10.1006/hbeh.1995.1277.
41. FLORES, D., A. TOUSIGNANT and D. CREWS (1994): Incubation temperature affects the behavior of adult leopard geckos (*Eublepharis macularius*). *Physiol. Behav.* 55, 1067-1072. 10.1016/0031-9384(94)90389-1.
42. GARDINER, D. W., F. M. BAINES and K. PANDHER (2009): Photodermatitis and Photokeratoconjunctivitis in a Ball Python (*Python regius*) and a Blue-Tongue Skink (*Tiliqua spp.*). *J. Zoo Wildl. Med.* 40, 757-766. 10.1638/2009-0007.1.
43. GÖRING, H. (2018): Vitamin D in Nature: A Product of Synthesis and/or Degradation of Cell Membrane Components. *Biochem.* 83, 1350-1357. 10.1134/S0006297918110056.
44. GOULD, A., L. MOLITOR, K. ROCKWELL, M. WATSON and M. A. MITCHELL (2018): Evaluating the Physiologic Effects of Short Duration Ultraviolet B Radiation Exposure in Leopard Geckos (*Eublepharis macularius*). *J. Herpetol. Med. Surg.* 28, 34. 10.5818/17-11-136.1.
45. HADDAD, J. G., L. HILLMAN and S. ROJANASATHIT (1976): Human serum binding capacity and affinity for 25-hydroxyergocalciferol and 25-hydroxycholecalciferol. *J. Clin. Endocrinol. Metab.* 43, 86-91. 10.1210/jcem-43-1-86.
46. HATHAWAY, J. A. and D. H. SLINEY (2016): Visible Light and Infrared Radiation. *Phys. Biol. Haz. Work*, 203-208. 10.1002/9781119276531.ch13.
47. HECKERS, K. O. and H. AUPPERLE (2014): Pigment-forming tumors in reptiles: light regime and its dark sides. *Ass. Rept. Amph. Vet.* 31-35.
48. HECKERS, K. O., H. AUPPERLE, V. SCHMIDT and M. PEES (2012): Melanophoromas and Iridophoromas in Reptiles. *J. Comp. Pathol.* 146, 258-268. 10.1016/j.jcpa.2011.07.003.
49. HOLICK, M. F., J. A. MACLAUGHLIN and S. H. DOPPELT (1981): Regulation of Cutaneous Previtamin D3 Photosynthesis in Man: Skin Pigment is not an Essential Regulator. *Am. Ass. Adv. Sci.* 211, 590-593.
50. HOLICK, M. F., X. Q. TIAN and M. ALLEN (1995): Evolutionary importance for the membrane enhancement of the production of vitamin D3 in the skin of poikilothermic animals. *Nat. Acad. Sci. USA.* 92, 3124-3126. 10.1073/pnas.92.8.3124.
51. HOLICK, M. (2004): Vitamin D: importance in the prevention of cancers, type 1 diabetes. *Am. J. Clin. Nutr.* 79, 362-371.
52. HOLICK, M. F. (2007): Vitamin D Deficiency. *New Eng. J. Med.* 357, 266-281. 10.1016/B978-1-4377-0987-2.00009-1.
53. HOLICK, M. F. (2014): How Much Sunlight Do We Need? In: Reicharth, J. (Ed.), *Sunlight, Ultraviolet Radiation, Vitamin D and Skin Cancer (2nd ed., pp. 1-16)*. Landes Biosci. Sp. Sci. Bus. Med.
54. HOLICK, M. F. (2018): Photobiology of Vitamin D. In: Feldman, D., J. W. Pike, R. Bouillon, E. Giovannucci, D. Goltzman, and M. Hewison (Eds.), *Vitamin D: Fourth Edition: Vol. 1: Biochem (4th ed., pp. 45-55)*. Elsevier Inc. 10.1016/B978-0-12-809965-0.00004-5.
55. HOLLIS, B. W. and C. L. WAGNER (2013): The role of the parent compound vitamin d with respect to metabolism and function: Why clinical dose intervals can affect clinical outcomes. *J. Clin. Endocrinol. Metab.* 98, 4619-4628. 10.1210/jc.2013-2653.
56. HONKAVAARA, J., M. KOIVULA, E. KORPIMAKI, H. SIITARI and J. VIITALA (2002): Ultraviolet vision and foraging in terrestrial vertebrates. *OIKOS* 98, 505-511.
57. HOSSEIN-NEZHAD, A. and M. F. HOLICK (2013): Vitamin D for health: A global perspective. *Mayo Clin. Proceed.* 88, 720-755. 10.1016/j.mayocp.2013.05.011.
58. HOW, K. L., H. A. W. HAZEWINDEL and J. A. MOL (1994): Dietary Vitamin D Dependence of Cat and Dog Due to Inadequate Cutaneous Synthesis of Vitamin D. *Gen. Comp. Endocrinol.* 96, 12-18.
59. HUANG, V. and D. CREWS (2012): Differences induced by incubation temperature, versus androgen manipulation, in male leopard geckos (*Eublepharis macularius*). *Physiol. Behav.* 107, 121-124. 10.1016/j.physbeh.2012.06.014.
60. HUANG, V., H. C. HEMMINGS and D. CREWS (2014): Sociosexual investigation in sexually experienced, hormonally manipulated male leopard geckos: Relation with phosphorylated DARPP-32 in dopaminergic pathways *J. Exp. Zool.* 321, 595-602. 10.1002/jez.1891.

61. IUCN, G. S. (n.d.). IUCN Red List of Threatened Species. <https://www.iucnredlist.org/search?query=leopard%20gecko&searchType=species>.
62. JABLONSKI, N. G. and G. CHAPLIN (2000): The evolution of human skin coloration. *J. Hum. Evol.* 39, 57-106. 10.1006/jhev.2000.0403.
63. JONES, G., D. E. PROSSER and M. KAUFMANN (2018): The Activating Enzymes of Vitamin D Metabolism (25- and 1 α -Hydroxylases). In: *Vitamin D: Fourth Edition: Vol. 1: Biochem (4th ed., pp. 57-79)*. Elsevier Inc. 10.1016/B978-0-12-809965-0.00005-7.
64. JUNQUEIRA, L. C. and J. CARNEIRO (2013): Pele e Anexos. In *Histologia Básica (12th ed., pp. 354-365)*. Guanabara Koogan Ltda.
65. JUZENIENE, A. and J. MOAN (2012): Beneficial effects of UV radiation other than via vitamin D production. *Dermatoendocrinol.* 4, 109-117. 10.4161/derm.20013.
66. KARSTEN, K. B., G. W. FERGUSON, T. CHEN and M. F. HOLICK (2009): Panther chameleons, *furcifer pardalis*, behaviorally regulate optimal exposure to UV depending on dietary vitamin D status. *Physiol. Biochem. Zool.* 82, 218-225. 10.1086/597525.
67. KHAN, M. S. (2016): Leopard gecko *Eublepharis macularius* from Pakistan Reptilia Natural history and biology of hobbyist choice leopard gecko *Eublepharis macularius* Muhammad Sharif Khan Herpetological Laboratory, January 2009.
68. KRATOCHVÍL, L. and D. FRYNTA (2002): Body size, male combat and the evolution of sexual dimorphism in eublepharid geckos (Squamata: Eublepharidae). *Biol. J. Linn. Soc.* 76, 303-314. 10.1046/j.1095-8312.2002.00064.x.
69. LARNER, D. P., J. S. ADAMS and M. HEWISON (2018): Regulation of Renal and Extrarenal 1 α -Hydroxylase. In: *Vitamin D: Fourth Edition: Vol. 1: Biochem (4th ed., pp. 117-137)*. Elsevier Inc. 10.1016/B978-0-12-809965-0.00008-2.
70. LI, H., M. J. VAUGHAN and R. K. BROWNE (2009): A complex enrichment diet improves growth and health in the endangered Wyoming Toad (*Bufo baxteri*). *Zoo Biol.* 28, 197-213. 10.1002/zoo.20223.
71. LOCK, B. (2017): Nutritional Secondary Hyperparathyroidism in Reptiles. *Vet. Inf. Network, Inc.* <https://veterinarypartner.vin.com/default.aspx?pid=19239&catId=102919&andId=8012396>.
72. LUPU, C. and S. ROBINS (2013): Determination of a Safe and Effective Ultraviolet B Radiant Dose in Budgerigars (*Melopsittacus undulatus*): A Pilot Study. *J. Avian Med. Surg.* 27, 269-279. 10.1647/2011-0291.
73. MANNING, B. and G. C. GRIGG (1997): Basking Is Not of Thermoregulatory Significance in the "Basking" Freshwater Turtle *Emydera signata*. *Copeia*, 1997(3), 579. 10.2307/1447562.
74. MARX, S. J., G. JONES, R. S. WEINSTEIN, G. P. CHROUSOS and D. M. RENQUIST (1989): Differences in mineral metabolism among nonhuman primates receiving diets with only vitamin D3 or only vitamin D2. *J. Clin. Endocrinol. Metab.* 69, 1282-1290. 10.1210/jcem-69-6-1282.
75. MCCOLLUM, E. V., N. SIMMONDS and J. ERNESTINE (1922): An Experimental Demonstration of the Existence of a Vitamin Which Promotes Calcium Deposition. *The J. Biol. Chem.* 53, 293-312.
76. MCDONALD, R. P. and M. K. VICKARYOUS (2018): Evidence for neurogenesis in the medial cortex of the leopard gecko, *Eublepharis macularius*. *Sci. Rep.* 8, 1-15. 10.1038/s41598-018-27880-6.
77. MCLEAN, K. E. and M. K. VICKARYOUS (2011): A novel amniote model of epimorphic regeneration: The leopard gecko, *Eublepharis macularius*. *BMC Dev. Biol.* 11. 10.1186/1471-213X-11-50.
78. MILES, S. (2017): Common conditions in leopard geckos (*Eublepharis macularius*). *Comp. Anim.* 22, 546-551. 10.12968/coan.2017.22.9.546.
79. MIRZA, Z. A., R. V. SANAP, D. RAJU, A. GAWAI and P. GHADKAR (2014): A new species of lizard of the genus *Eublepharis* (Squamata: Eublepharidae) from India. *Phyllomed.* 13, 75-90. 10.11606/issn.2316-9079.v13i2p75-90.
80. NAGPAL, S., S. NA and R. RATHNACHALAM (2005): Noncalcemic actions of vitamin D receptor ligands. *Endocr. Rev.* 26, 662-687. 10.1210/er.2004-0002.
81. NAKAMICHI, Y., N. TAKAHASHI, N. UDAGAWA and T. SUDA (2018): Osteoclastogenesis and Vitamin D. In: Feldman, D., J. W. Pike, R. Bouillon, E. Giovannucci, D. Goltzman, and M. Hewison (Eds.), *Vitamin D: Fourth Edition: Vol. 1: Biochem (4th ed., pp. 309-317)*. Elsevier Inc. 10.1016/B978-0-12-809965-0.00018-5
82. NAKASHIMA, N. (2016): Regeneration of dermal patterns from the remaining pigments after surgery in *Eublepharis macularius* (a case report). *BMC Vet. Res.* 12, 10-13. 10.1186/s12917-016-0765-x.
83. O'MALLEY, B. (2005): General anatomy and physiology of reptiles. In: O'Malley, B. (Ed.), *Clinical Anatomy and Physiology of Exotic Species (1st ed., pp. 17-39)*. Elsevier Ltd.
84. O'MALLEY, B. (2008): *Nutritional Problems in Reptiles*. WSAVA.
85. OONINCX, D. G. A. B., Y. STEVENS, J. J. G. C. VAN DEN BORNE, J. P. T. M. VAN LEEUWEN and W. H. HENDRIKS (2010): Effects of vitamin D3 supplementation and UVB exposure on the growth and plasma concentration of vitamin D3 metabolites in juvenile bearded dragons (*Pogona vitticeps*). *Comp. Biochem. Physiol.* 156, 122-128. 10.1016/j.cbpb.2010.02.008.
86. OREN, M. and J. BARTEK (2007): The Sunny Side of p53. *Cell* 128, 826-828. 10.1016/j.cell.2007.02.027.
87. PANSU, D., C. BELLATON and F. BRONNER (1981): Effect of Ca intake on saturable and nonsaturable components of duodenal Ca transport. *Am. J. Physiol.* 3, 32-37. 10.1152/ajpgi.1981.240.1.g32.
88. PANSU, D., C. BELLATON, C. ROCHE and F. BRONNER (1983): Duodenal and ileal calcium absorption in the rat and effects of vitamin D. *Am. J. Physiol.* 7. 10.1152/ajpgi.1983.244.6.g695.

89. PEACOCK, H. M., E. A. B. GILBERT and M. K. VICKARYOUS (2015): Scar-free cutaneous wound healing in the leopard gecko, *Eublepharis macularius*. *J. Anat.* 227, 596-610. 10.1111/joa.12368.
90. PENNING, M. R. (2012): Diet Supplementation in Reptiles: Prevention and Treatment of Common Disorders. NAVC Conference 2012 Small Animal.
91. POKORNÁ, M., M. RÁBOVÁ, P. RÁB, M. A. FERGUSON-SMITH, W. RENS and L. KRATOCHVÍL (2010): Differentiation of sex chromosomes and karyotypic evolution in the eyelid geckos (Squamata: Gekkota: Eublepharidae), a group with different modes of sex determination. *Chromosome Res.* 18, 809-820. 10.1007/s10577-010-9154-7.
92. POLZONETTI, V., S. PUCCIARELLI, S. VINCENZETTI and P. POLIDORI (2020): Dietary Intake of Vitamin D from Dairy Products Reduces the Risk of Osteoporosis. *Nutrients* 12, 1-15. 10.3390/nu12061743.
93. PUTZ, O. and D. CREWS (2005): Embryonic origin of mate choice in a lizard with temperature-dependent sex determination. *Dev. Psychobiol.* 48, 29-38. 10.1002/dev.20109.
94. RAITI, P. and M. M. GARNER (2006): Metastatic Mineralization in a Geoffrey's Side-Necked Turtle, *Phrynops geoffroanus*. *J. Herpetol. Med. Surg.* 16, 135-139. 10.5818/1529-9651.16.4.135.
95. RAWAT, Y. B., K. BAHADUR THAPA, S. BHATTARAI and K. B. SHAH (2019): First Records of the Common Leopard Gecko, *Eublepharis macularius* (Blyth 1854) (Eublepharidae), in Nepal. *Reptiles and Amphibians* 26, 89.
96. RHEN, T. and D. CREWS (2000): Organization and activation of sexual and agonistic behavior in the leopard gecko, *Eublepharis macularius*. *Neuroendocrinol.* 71, 252-261. 10.1159/000054543.
97. ROELANDTS, R. (2002): The history of phototherapy: Something new under the sun? *J. Am. Acad. Dermatol.* 46, 926-930. 10.1067/mjd.2002.121354.
98. ROSSI, J. V. (2019). General Husbandry and Management. In: Divers, S. J. and S. J. Stahl (Eds.), *Mader's Reptile and Amphibian Medicine and Surgery* (3rd ed., pp. 109-129).
99. RUTLAND, C. S., P. CIGLER and V. KUBALE (2019): Reptilian Skin and Its Special Histological Structures. In: *Veterinary Anatomy and Physiology*. IntechOpen. 10.5772/intechopen.84212.
100. SCOTT, G. N., H. H. NOLLENS and T. L. SCHMITT (2019): Evaluation of Plasma 25-Hydroxyvitamin D, Ionized Calcium, and Parathyroid Hormone in Green Sea Turtles (*Chelonia mydas*) Exposed to Different Intensities of Ultraviolet B Radiation. *Am. Ass. Zoo Vet.* 50, 421-426.
101. SLAVENS, F. and K. SLAVENS (2001): Reptiles and amphibians in captivity: longevity home page.
102. STANFORD, M. (2006): Effects of UVB radiation on calcium metabolism in psittacine birds. *Vet. Rec.* 159, 236-241. 10.1136/vr.159.8.236.
103. STEENBOCK, H., A. BLACK, M. T. NELSON, C. A. HOPPERT and B. M. RIISING (1925): The induction of growth-promoting and calcifying properties in fats and their usaponifiable constituents by exposure to light. *J. Biol. Chem.* 64, 263-298.
104. STRZELEWICZ, M. A., D. E. ULLREY, S. F. SCHAFER and J. P. BACON (1985): Feeding Insectivores: Increasing the Calcium Content of Wax Moth (*Galleria mellonella*) Larvae. *J. Zoo Anim. Med.* 16, 25. 10.2307/20094728.
105. SUBRAMANIAM, N., J. J. PETRIK and M. K. VICKARYOUS (2018): VEGF, FGF-2 and TGF β expression in the normal and regenerating epidermis of geckos: implications for epidermal homeostasis and wound healing in reptiles. *J. Anat.* 232, 768-782. 10.1111/joa.12784.
106. SUDA, T., R. MASUYAMA, R. BOUILLON and G. CARMELIET (2015): Physiological functions of vitamin D: what we have learned from global and conditional VDR knockout mouse studies. *Curr. Opin. Pharmacol.* 22, 87-99. 10.1016/j.coph.2015.04.001.
107. TANAKA, Y. and H. F. DELUCA (1981): Measurement of mammalian 25-hydroxyvitamin D3 24R- and 1 α -hydroxylase. *Proc. Natl. Acad. Sci. USA.* 78, 196-199. 10.1073/pnas.78.1.196.
108. TAYLOR, S. L., M. KAUR, K. LOSICCO, J. WILLARD, F. CAMACHO, K. S. O'ROURKE and S. R. FELDMAN (2009): Pilot study of the effect of ultraviolet light on pain and mood in fibromyalgia syndrome. *J. Altern. Complement. Med.* 15, 15-23. 10.1089/acm.2008.0167.
109. SZYDŁOWSKI, P., J. P. MADEJ and M. MAZURKIEWICZ-KANIA (2015): Ultrastructure and distribution of chromatophores in the skin of the leopard gecko (*Eublepharis macularius*). *Acta Zool.* 97, 370-375. 10.1111/azo.12132.
110. VELEVA, B. I., R. L. VAN BEZOOIJEN, V. G. M. CHEL, M. E. NUMANS and M. A. A. CALJOUW (2018): Effect of ultraviolet light on mood, depressive disorders and well-being. *Photodermatol. Photoimmunol. Photomed.* 34, 288-297. 10.1111/phpp.12396.
111. VERGNEAU-GROSSET, C. and F. PÉRON (2020): Effect of ultraviolet radiation on vertebrate animals: update from ethological and medical perspectives. *Photochem. Photobiol. Sci.* 19, 752-762. 10.1039/c9pp00488b.
112. VIETS, B. E., A. TOUSIGNANT, M. A. EWERT, C. E. NELSON and D. CREWS (1993): Temperature-dependent sex determination in the leopard gecko, *Eublepharis macularius*. *J. Exp. Zool.* 265, 679-683. 10.1002/jez.1402650610.
113. WACKER, M. and M. F. HOLICK (2013a): Sunlight and Vitamin D: A global perspective for health. *Dermatoendocrinol.* 5, 51-108. 10.4161/derm.24494.
114. WACKER, M. and M. F. HOLICK (2013b): Vitamin D-effects on skeletal and extraskeletal health and the need for supplementation. *Nutrients* 5, 111-148. 10.3390/nu5010111.

115. WALLACH, J. D. (1966): Hypervitaminosis D in green iguanas. *J. Am. Vet. Med. Assoc.* 149, 912-914.
116. WANGEN, K., J. KIRSHENBAUM and M. A. MITCHELL (2013): Measuring 25-Hydroxy Vitamin D Levels in Leopard Geckos exposed to Commercial Ultraviolet B Lights. *Ass. Rept. Amph. Vet.* 42.
117. WATSON, M. K., A. W. STERN, A. L. LABELLE, S. JOSLYN, T. M. FAN, K. LEISTER, M. KOHLES, K. MARSHALL and M. A. MITCHELL (2014): Evaluating the clinical and physiological effects of long term ultraviolet B radiation on guinea pigs (*Cavia porcellus*). *PLoS ONE* 9, 1-23. 10.1371/journal.pone.0114413.
118. WATSON, M. K. and M. A. MITCHELL (2014): Vitamin D and Ultraviolet B Radiation Considerations for Exotic Pets. *J. Exot. Pet Med.* 23, 369-379. 10.1053/j.jepm.2014.08.002
119. WEBB, A. R., B. R. DECOSTA and M. F. HOLICK (1989): Sunlight regulates the cutaneous production of vitamin D3 by causing its photodegradation. *J. Clin. Endocrinol. Metab.* 68, 882-887. 10.1210/jcem-68-5-882.
120. WEBB, A. R. and M. F. HOLICK (1988): The Role Of Sunlight In The Cutaneous Production Of Vitamin D3. *Annu. Rev. Nutr.* 8, 375-399. 10.1146/annurev.nutr.8.1.375.
121. WILKINSON, S. L. (2015): Reptile wellness management. *Vet. Clin. North Am. Exot. Anim. Pract.* 18, 281-304. 10.1016/j.cvex.2015.01.001.
122. ZEHNDER, D., R. BLAND, M. C. WILLIAMS, R. W. MCNINCH, A. J. HOWIE, P. M. STEWART and M. HEWISON (2001): Extrarenal Expression of 25-Hydroxyvitamin D 3 -1 α -Hydroxylase 1. *J. Clin. Endocrinol. Metabol.* 86, 888-894. 10.1210/jcem.86.2.7220.
123. ZEHNDER, D., R. BLAND, R. S. CHANA, D. C. WHEELER, A. J. HOWIE, M. C. WILLIAMS, P. M. STEWART and M. HEWISON (2002): Synthesis of 1,25-dihydroxyvitamin D3 by human endothelial cells is regulated by inflammatory cytokines: A novel autocrine determinant of vascular cell adhesion. *J. Am. Soc. Nephrol.* 13, 621-629.
124. ZWART, P. and R. J. RULKENS (1979): Improving the calcium content of mealworms. *Int. Zoo Yearb.* 19, 254-255. 10.1111/j.1748-1090.1979.tb00574.x.

Pregled stila života leopard gekona i važnost ultraljubičastog zračenja, vitamina D i kalcija

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Vrstu leopard gekon (*Eublepharis macularius*) prvi put je opisao 1854. britanski herpetolog Edward Blyth. Danas je to jedna od najpopularnijih vrsta guštera za kućnog ljubimca, uglavnom zbog njegovog mirnog temperamenta, jednostavnosti održavanja i reprodukcije u zatočeništvu, njegove dugovječnosti i male veličine te njegove ljepote i raznolikosti boja i uzoraka. Ultraljubičasto (UV) zračenje je važan regulator ponašanja.

Vitamin D regulira i brojne osnovne fiziološke funkcije u kralježnjaka, uglavnom homeostazu kalcija. Zbog velike raznolikosti vrsta gmazova i različitih prilagodbi okolišu, važno je znati potrebe i prilagodbe svake vrste u svezi UV, vitamina D i kalcija. Ovaj rad pruža čitateljima pregled stila života leopard gekona i važnosti ultraljubičastog zračenja, vitamina D i kalcija za tu vrstu.

Ključne riječi: ponašanje, *Eublepharis macularius*, habitat, život, dodatak prehrani